

RADIATION-INDUCED HEMATOLOGICAL ALTERATIONS AND THEIR INHIBITION BY *AEGLE* *MARMELOS* FRUIT EXTRACT

by

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This study was carried out to observe the radio protective potential of *Aegle Marmelos* fruit extract (AME) against radiation-induced hematological and biochemical alterations in blood and liver of mice. For this purpose, adult Swiss albino mice were exposed to 6 Gy gamma radiation in the presence (experimental) or absence (control) of the extract (100 mg/kg body weight animal/day). Exposure to radiation resulted in a significant decline in the count of erythrocyte, hemoglobin (Hb) and hematocrit (Hct) in peripheral blood. In contrast, extract-pretreated irradiated animals had a significant rise in all of these blood constituents, as compared with the irradiated control. Furthermore, a significant elevation in lipid peroxidation over normal was recorded in the irradiated control, whereas such increase was considerably lesser in extract-pretreated animals. Likewise, pretreatment with AME caused a significant increase in glutathione levels in the serum, as well as in the liver, in comparison to irradiated controls. These results indicate that AME may be responsible for the protection of stem cells in bone marrow, subsequently resulting in a rise of hematological constituents in peripheral blood. The present study affirms the prophylactic use of AME against radiation-induced hematological and biochemical alterations in mammals.

Key words: gamma radiation, glutathione, hematological constituents, lipid peroxidation, *Aegle Marmelos*, Swiss albino mice

INTRODUCTION

Radiation is the most studied environmental hazard in the world. Ionizing radiations including alpha, beta, and gamma rays and neutrons with sufficient energy to generate ion pairs, *i. e.* electrons which can generate chemically active free radicals can, in turn, damage the molecular structure, resulting in cellular dysfunctions or mutations [1]. The inadvertent exposure of humans to various sources of radiation causes the ionization of molecules, setting off potentially damaging reactions, via free radicals production. Free radicals are believed to play a role in more than sixty different health conditions, including the ageing process, cancer, radiation damage, atherosclerosis *etc.* [2, 3].

In today's highly nuclear threatened environment, there is an increased need to protect not only high risk service groups from the hazards of unintended ionizing radiation exposure, but the general

public, as well. Thus, there is an urgent need for the development of radioprotective agents. Fortunately, there are many plant derived natural antioxidants that interfere with free radicals before they can damage the body. Antioxidants work in several ways, either by reducing the energy of the free radicals, stopping the free radicals from forming in the first place, or interrupting an oxidizing chain reaction and, thus, minimizing the damage this causes.

Radiation damage results from the sensitivity of cells to radiation, with those that replicate most rapidly being those most sensitive to radiation exposure. Mature cells that are more highly differentiated appear to be least affected by radiation. This difference in cell sensitivity is the basis for the distinction among the three sub-syndromes of the acute radiation syndrome (ARS). ARS is divided into hematopoietic, gastro-intestinal, and neurovascular sub-syndromes. Human beings overexposed to radiations are prone to developing life-threatening diseases, often related to the hematopoietic system. This being the result of the fact that the hematopoietic system is highly sensitive to ra-

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diation and that peripheral blood counts may well serve as a biological indicator of such damage. The target cells of the hematopoietic tissues are the stem cells.

Radiation protection concepts and philosophy have been evolving over the past several decades. Several synthetic compounds have been used in the past as radio protectors. For the first time, Patt *et al.*, observed that the pretreatment of rats and mice with cysteine (naturally occurring amino acid) before exposure to radiation protected them against radiation-induced sickness and mortality [4]. Subsequently, several chemical compounds were synthesized and tested for their radioprotective ability [5]. However, the major drawback of these compounds has been their high toxicity at optimum doses. Therefore, there is a need to screen alternatives which are non-toxic or less toxic at their optimum protective doses for practical purposes.

Herbal drugs offer an alternative to synthetic compounds and have been considered either non-toxic or less toxic, thus giving an impetus to the screening of their radio protective properties. *Aegle Marmelos*, commonly known as bael, is a spinous tree belonging to the family *Rutaceae*. It is widely found in India, Bangladesh, Burma, and Sri Lanka. It is distributed mainly within the sub-Himalayan forests, in dry hilly regions. It is called Shivadume, the tree of lord Shiva. *Aegle Marmelos* plays an important role in the indigenous system of Indian medicine. Its edible leaf, root, bark, seed, and fruit are highly valued in Ayurvedic medicine in India [6]. In fact, since Charaka (1500 BC), no drug has been longer or better known or appreciated by the inhabitants of India than the bael [7]. The fruit of this plant has been used as an astringent, relief for indigestion, and cure for the treatment of diarrhea, dysentery, and stomachalgia. Aqueous AME exhibits an anti-hyperlipidaemic [8] and hypoglycemic [9] effect in ptozotocin-induced diabetic rats. The ripe fruit of this plant is used in different formulae for the treatment of chronic diarrhea [10].

Envisioning a possible use of the pharmacological and therapeutic values of this plant, the present study has been carried out in order to access the protective potential of the fruit of *Aegle Marmelos* against radiation-induced hematological and biochemical alterations in mammals.

MATERIALS AND METHODS

Animal care and handling

Animal care and handling were carried out according to the guidelines set by the World Health Organization, Geneva, and the Indian National Science Academy (INSA), New Delhi. Male Swiss albino mice (*Mus Musculus*), 6 to 8 weeks old and weighing 20-24 g, from an inbred colony, were used for the present study. These animals were kept under controlled conditions of temperature and light (light: dark, 10-14

hours). They were provided with standard mice feed (procured from Aashirwad Industries, Chandigarh, India) and water ad libitum. As a preventive measure against infections, tetracycline water was given once a fortnight. This study has the approval of the Departmental Animal Ethical Committee.

Irradiation

The Cobalt Teletherapy Unit (ATC-C9) at the Cancer Treatment Centre, Department of Radiotherapy, SMS Medical College & Hospital, Jaipur, India, was used for irradiation. Unanesthetized mice were restrained in a well ventilated Perspex box and whole-body exposed to 6 Gy gamma radiation.

Preparation of *Aegle Marmelos* extract

The fruits of *Aegle Marmelos* L. were collected locally after their proper identification by a competent botanist (Voucher Specimen no: RUBL-20438) from the herbarium, Department of Botany, University of Rajasthan, Jaipur, Rajasthan, India. The pulp was removed from the fruit and shade dried, powdered into a mixture and its hydro-alcoholic extract then prepared by refluxing it with double-distilled water (DDW) and alcohol (3:1) for 36 hours (12 + 3 hours) at 40 °C. The liquid extract was cooled and concentrated by evaporating its liquid content. The prepared AME was stored at a low temperature until further use. The extract was re-dissolved in DDW prior to oral administration to mice.

EXPERIMENTAL DESIGN

Determination of the optimum dose of AME against irradiation

Different doses of AME were tested against 8 Gy gamma radiations in Swiss albino mice in order to find out the optimal dose of AME on the basis of the survival percentage of such animals after up to 30 days of irradiation.

Modification of radiation response

A total of 70 animals used in the experiment were assorted into 4 groups. Five animals in Group I were administered with DDW, volume equal to AME (100 mg/kg body weight animal/day) by oral gavage, to serve as normal (vehicle treated); five mice in Group II were administered AME orally, once a day, with a dose of 100 mg/kg body weight animal/day for 5 consecutive days. In Group III, DDW volume equal to AME was administered for 5 consecutive days (as in

Group I). One hours after the last administration of DDW, such animals ($n = 30$) were exposed to 6 Gy gamma rays. Group IV mice ($n = 30$) were treated with AME orally for 5 consecutive days (as in Group II) and exposed to gamma radiation 1 hour after the last administration of AME on the 5th day.

All animals were observed daily for any signs of sickness, morbidity, behavioral toxicity, mortality and abnormality, if any. A minimum of 5 animals from each group were necropsied at 12 hours on days 1st, 3rd, 7th, 15th, and 30th post-treatment, in order to evaluate hematological and biochemical alterations.

Hematological study

Blood was collected from the orbital sinuses of animals from each group in a vial containing 0.5 M EDTA (ethylene diamine tetra acetic acid). Total numbers of erythrocyte (RBC), hematocrit (Hct), and hemoglobin (Hb) content were determined using standard procedures.

Biochemical study

Lipid peroxidation (LPO) assay: The lipid peroxidation (LPO) level in liver and blood serum was measured in terms of thiobarbituric acid reactive substances (TBARS) after 24 hours, using the method of Okhawa *et al.* [11]. The absorbance was read at 532 nm, using a UV-VIS Systronics spectrophotometer.

Glutathione assay: The hepatic level of reduced glutathione (GSH) was determined after 24 hours by the Moron *et al.* [12] method. The GSH content in blood was measured spectrophotometrically, using Ellman's reagent with 5-5, dithiobis-2-nitrobenzoic acid (DTNB) as a coloring reagent, according to the method of Beutler *et al.* [13]. The absorbance was read at 412 nm.

Statistical analysis

The results for all groups at various necropsy intervals were expressed as a mean standard error (S. E.). Statistical differences between various groups were analyzed by the Student's t test and their significance observed at different levels as $p < 0.05$, $p < 0.01$, and $p < 0.001$.

RESULTS

Radiation sickness and mortality

No toxic effects in terms of sickness were observed in the animals treated with DDW (Group I) and AME alone (Group II). Some of the animals exhibited signs of radiation sickness, such as anorexia, lethargy, diarrhea, body weight loss, and ruffled fur within 4 days after radiation exposure (Group III). No adverse effects were observed in terms of sickness, body weight, urination, defecation pattern, and mortality in the irradiated animals (Group IV).

Selection of the optimum dose of AME against irradiation

The optimum dose of AME against lethal gamma radiation (*i. e.* 8 Gy) for Swiss albino mice was selected on the basis of the survival experiment, where a number of deaths and survivals were recorded for up to 30 days after irradiation. Mice treated with AME, at doses of 25, 50, 100, 200, 400, and 800 mg/kg body weight day for 5 consecutive days prior to irradiation, exhibited a 30%, 55%, 88%, 62%, 42%, and 33% rate of survival, respectively. The dose of 100 mg/kg body weight was found to be the optimum dose based on the above data and further studies were carried out using this dose of AME (fig. 1).

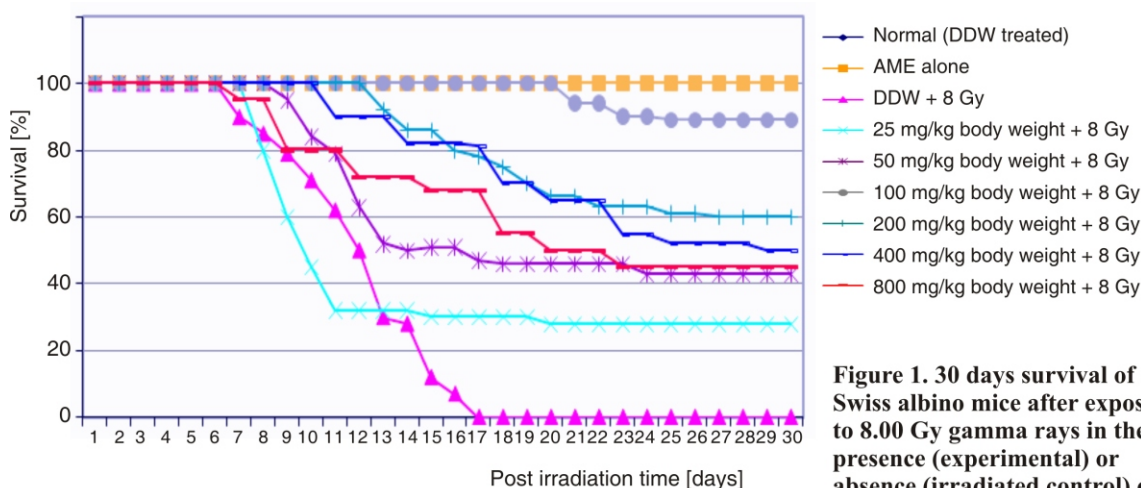


Figure 1. 30 days survival of Swiss albino mice after exposure to 8.00 Gy gamma rays in the presence (experimental) or absence (irradiated control) of AME

Hematological study

Animals treated with AME alone (Group II) did not show any significant changes in various hematological constituents (erythrocytes, Hb & Hct) in comparison with the normal (Group I). Throughout the experiment, erythrocytes showed a significant decrease from normal. The maximum decrease in the total erythrocyte count was scored on day 3rd, after which such cells increased in their number on day 7th, but the normal value could not be restored even at the last autopsy interval (*i. e.* day 30). A significant increase in the red cell count was noticed during the entire period of the study, returning to an almost normal value at the last autopsy interval (*i. e.* day 30; fig. 2).

Hemoglobin concentration in irradiated mice showed the maximum decrease on day 3rd. Later, a slight increase was observed from day 7th on, but the values remained below normal up to the last autopsy interval. Animals irradiated with pretreated AME ex-

hibited a higher Hb concentration than Group III animals and values were found to be near normal by the end of the experiment (fig. 3).

Hematocrit percentage was found to be significantly lower in the irradiated control mice, with a maximum decline on day 3rd, recovery from day 7th on, and a return to near normal by the 30th day of irradiation. On the contrary, AME pretreated irradiated animals exhibited significantly higher hematocrit values throughout the experiment (fig. 4).

Lipid peroxidation assay

No significant difference in the levels of LPO in blood or liver was noticed between the sham-irradiated (Group I) and AME alone treated (Group II) animals. A significant increase in blood and hepatic LPO levels was noted in gamma-irradiated animals (Group III), as compared with normal animals. However, these levels were found to be significantly lower than in the AME-pretreated irradiated (Group IV) animals (fig. 5).

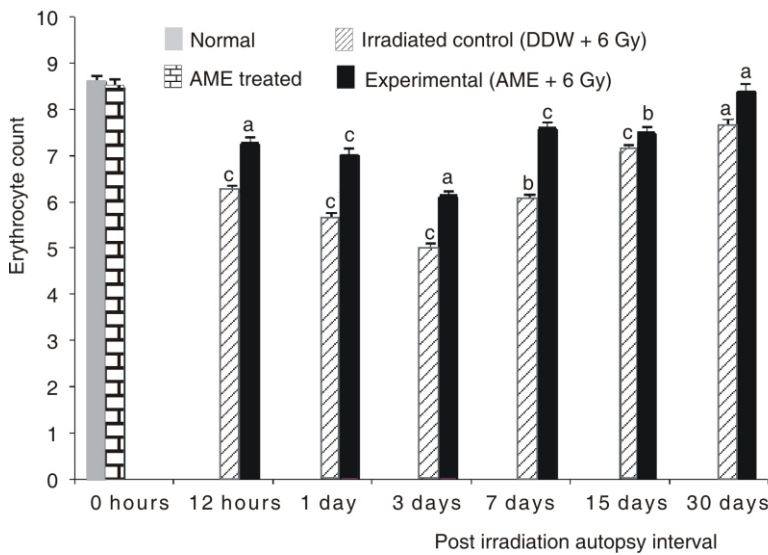


Figure 2. Variations (mean ± S. E.) in erythrocyte count (10⁶/mm³) in peripheral blood of Swiss albino mice after exposure to gamma rays with (experimental) or without (irradiated control) of AME
 Significance level: normal vs. irradiated control; irradiated control vs. experimental p values: (a) 0.05 b) 0.01, (c) 0.001

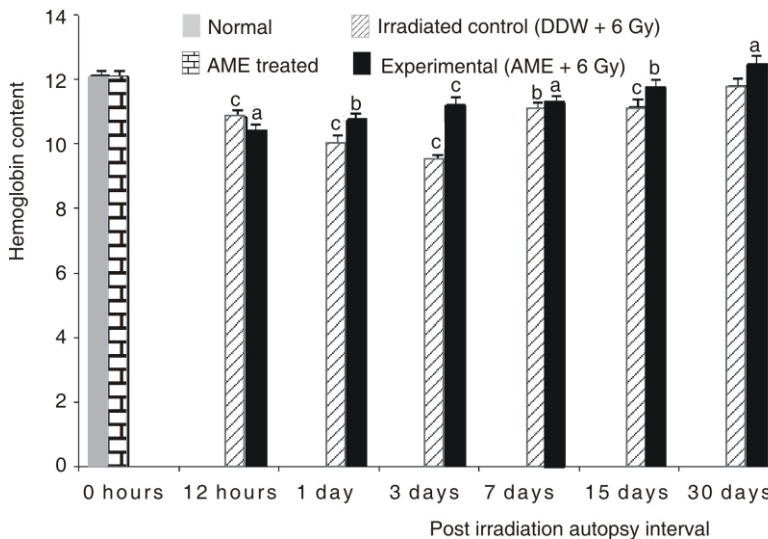


Figure 3. Variations (mean ± S. E.) in hemoglobin content (gm/dl) in peripheral blood of Swiss albino mice after exposure to gamma rays with (experimental) or without (irradiated control) of AME
 Significance level: normal vs. irradiated control; irradiated control vs. experimental p values: (a) 0.05 b) 0.01, (c) 0.001

Figure 4. Variations (mean ± S. E.) in hematocrit [%] in peripheral blood of mice after exposure to gamma rays with (experimental) or without (irradiated control) of AME

Significance level: normal vs. irradiated control; irradiated control vs. experimental
 p values: (a) ≤0.05, (b) ≤0.01, (c) ≤0.001

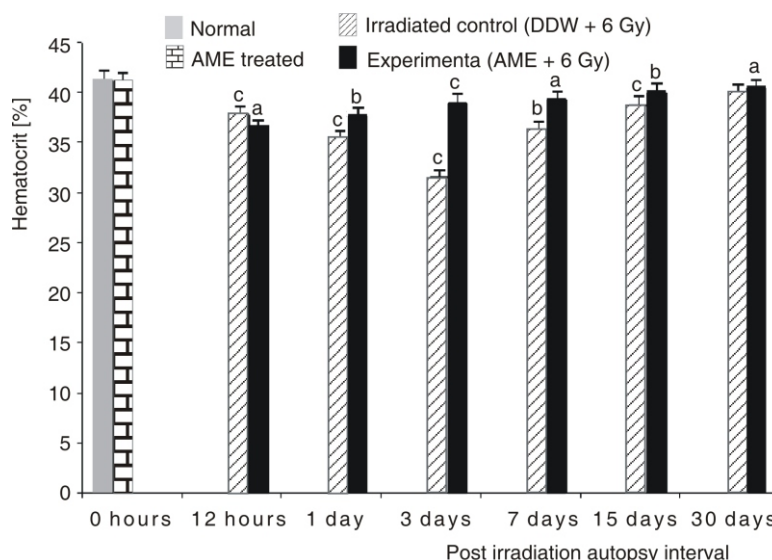
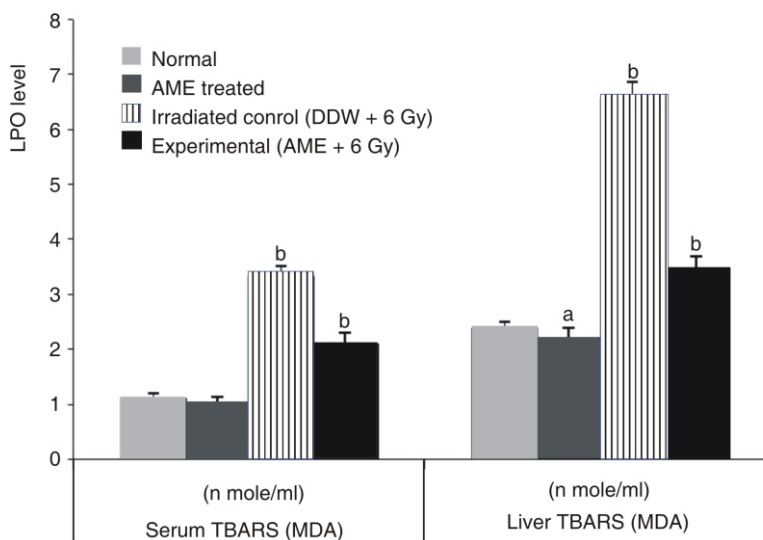


Figure 5. Lipid peroxidation level in terms of TBARS (MDA) in blood and liver of Swiss albino mice after exposure to 6 Gy of gamma rays with (experimental) or without (irradiated control) of AME

Significance level: normal vs. irradiated control; irradiated control vs. experimental
 p value: (a) 0.05 b ≤0.01



Glutathione estimation

No significant alterations in GSH contents of either liver or blood were observed between normal and AME-treated animals. However, a statistically significant decrease in GSH was noted in the irradiated control (Group III) in comparison with Group I animals. The AME-pretreated irradiated (Group IV) animals exhibited a significant elevation in GSH, both in blood and liver, in comparison with Group III animals; however, the values remained below normal (fig. 6).

DISCUSSION

The mortality of animals after irradiation in the present study may be due to the hematopoietic syndrome. Such deaths can be correlated with the impairment of the immune system [14]. Endogenous infections may also be responsible for the deaths of irradiated mice. Bacteremia may be one of the causes

of mortality secondary to hematopoietic and gastro-intestinal radiation damage, because antibiotic treatment has shown to increase the survival of mice irradiated in the LD_{50/30} range [15, 16].

The radioprotective effect of AME has been demonstrated by the increased survival rate. Significant radioprotection was achieved when AME was given orally in doses of 100 mg/kg body weight day for 5 consecutive days prior to irradiation. In our experiment, a 12% of mortality was evident at the said dose of AME. Similarly, plants such as *Emblica Officinalis* [17], *Rosemarinus Officinalis* [18] and *Alstonia Scholaris* [19] have also been reported to provide protection against radiation-induced sickness.

In the present study, signs of sickness and mortality were not observed in the sham-irradiated (Group-I) or AME alone treated (Group-II) animals. Within it, body weight loss in irradiated animals may be attributed to the reduced food and water intake, fluid loss by diarrhea and the diminished absorption capacity of the GI tract. In general, the present findings

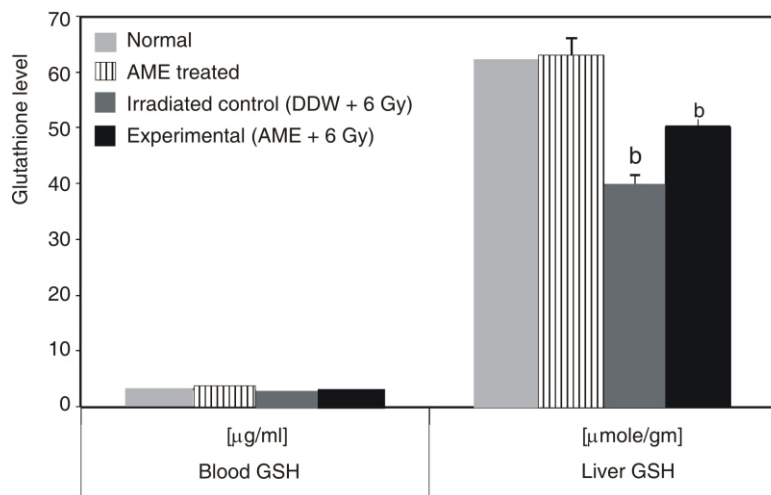


Figure 6. Reduced glutathione (GSH) level in blood and liver of Swiss albino mice after exposure to 6 Gy gamma rays with (experimental) or without (irradiated control) of AME

Significance level: normal vs. irradiated control; irradiated control vs. experimental p values: (a) ≤ 0.05 , (b) ≤ 0.01

are in agreement with those of Jagetia *et al.* [20], and Chaudhary *et al.* [21], with the exception of doses of gamma radiation in mammals.

The hematopoietic system is one of the most radiosensitive systems and damage done to it may play a crucial role in the development of the hematopoietic syndrome, possibly resulting in death. The sensitivity of blood cells in man to radiation determines the order in which the drop in the respective counts occurs [22]. Radiation-induced hematological changes in peripheral blood have been extensively studied, owing to their physiological significance [23, 24].

In the present study, the reduction in blood components in the irradiated group may be attributed to the impairment of cell division, obliteration of blood-forming organs, alimentary tract injury [25], depletion of factors needed for erythroblast differentiation and reticulocyte release from the bone marrow [26], as well as to the loss of cells from the circulation by hemorrhage or leakage through capillary walls and/or the direct destruction of mature circulating cells [27]. The decrease in all these blood constituents is responsible for anemia. Maximum decline in erythrocytes, hemoglobin, and hematocrit was observed on the third day following irradiation, which is in agreement with the findings in earlier works [18, 28, 29].

AME pretreatment before irradiation significantly checked the radiation-induced decline in erythrocytes, Hb and Hct. AME may be responsible for a significant protection of erythropoietic cells in bone marrow, which is subsequently responsible for the increase in such hematological components. Bone marrow cells have been reported to be protected against radiation-induced damage by various other plant extracts, as well [20, 30, 31].

The products of lipid peroxidation such as malonaldehyde and 4-hydroxynonenal are toxic to living cells [16, 32]. In the present study, it has been observed that although AME treatment did not significantly alter the LPO level in non-irradiated animals, it

significantly lowered the radiation-induced LPO in terms of malondialdehyde formation. Inhibition of LPO in biomembranes can be attributed to various antioxidants present in the extract of this plant. Similarly, other plant extracts containing antioxidants have been found conducive to checking the radiation-induced elevation of lipid peroxidation levels in mammal blood and liver [19, 21].

GSH is a versatile protector and executes its radioprotective function through free radical scavenging, restoration of damaged molecules by hydrogen donation, reduction of peroxides and maintenance of protein thiols in a reduced state [33]. A significant decline in GSH content from normal, in blood and liver both, was noticed after irradiation. This could be because of its enhanced utilization as an attempt to detoxify the free radicals generated by radiation. Oral administration of AME did not influence the endogenous GSH content significantly, but it protected against GSH depletion caused by irradiation. These results suggest that endogenous non-protein sulfhydryl content (GSH) is maintained by the extract in the AME-irradiated group. Various plant products have been reported earlier as inhibitors of radiation-inducing the decline of GSH content in various tissues in mammals [17-19, 21].

Natural antioxidants exhibit a long window of protection, *i. e.*, they provide some protection when administered hours before radiation exposure. Exogenous administration of antioxidants, such as glutathione, superoxide dismutase (SOD), antioxidant vitamins (A, C, and E), lipoic acid, as well as substances that mimic or induce the activity of endogenous antioxidant systems (*e. g.*, selenium, zinc, copper salts, and metal complexes), have exhibited protective properties against hematopoietic syndrome death [34-37]. Several phytochemical constituents like aegelin, alloimperatorin, marmelide, marmeline, marmelosin, marmesin, psoralen, skimming, tannic acid, xanthotoxol and β -sitosterol are reported to be present in the *Aegle*

Marmelos fruit [38-40]. The phytochemicals present in this plant might be responsible for the reduction of radiation-induced lipid peroxidation and the protection of erythropoietic stem cells in bone marrow, subsequently resulting in the increased levels of various hematological components in the peripheral blood noted in the present study. Since significant protection is obtained with a non toxic dose, AME may well have an advantage over known radioprotectors. Further research in order to determine the exact mechanism and clinical applicability of *Aegle marmelos* as a radioprotector is in progress at the moment.

CONCLUSION

Present results indicate that the use of *Aegle Marmelos* fruit extract possibly inhibits radiation-induced hematological alterations and oxidative stress during planned and unplanned radiation exposure in mammals.

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REFERENCES

- [1] Masumura, K., et al., Ion-Induced Mutations in the gpt Delta Transgenic Mouse: Comparison of Mutation Spectra Induced by Heavy-Ion, X-ray, and Gamma-Ray Radiation, *Environ Mol Mutagen*, 40 (2002), 3, pp. 207-215
- [2] Ames, B. N., Shigenaga, M. K., Hagen, T. M., Oxidants, Antioxidants, and the Degenerative Diseases of aging, USA, *Proc. Natl. Acad. Sci.*, 90 (1993), pp. 7915-7922
- [3] La Verne, J. A., OH Radicals and Oxidizing Products in the Gamma Radiolysis of Water, *Radiat. Res.*, 153 (2000), pp. 196-200
- [4] Patt, H. M., et al., Cysteine Protection Against X-Irradiation, *Science*, (1949), 110, pp. 213-214
- [5] Sweeny, T. R., Survey of Compounds from the Antiradiation Drug Development Program of the U. S. Army Medical Research and Development Command, Government Printing Office, Washington D. C. Publication, 1979, pp. 308-318
- [6] Sharma, R. K., Bhagwan, D., Agnivesa's Charaka Samhita, Chaukhamba Orientalia, Varanasi, 3, 1988
- [7] ***, CHEMEXCIL Selected Medicinal Plant of India, Basic Chemicals, Pharmaceutical and Cosmetic Export Promotion Council, Bombay, 1992, pp. 205-207
- [8] Miyazaki, S., Ranganathan, D., Kadarkarismami, M., Elucidation of Toxicity of the *A. Marmelos*, *Phytomedicine*, (2007), 4, pp. 204-205
- [9] Kamalakkannan, N., Rajadurai, M., Prince, P. S., Effect of *Aegle Marmelos* Fruits on Normal and Streptozotocin-Diabetic Wistar Rats., *J. Med Food*, (2003), 6, pp. 93-98
- [10] Citarasu, T., et al., Influence of Selected Indian Immunostimulant Herbs against white Spot Syndrome Virus (WSSV) Infection in Black Tiger Shrimp, *Penaeus Monodon* with Reference to Haematological, Biochemical and Immunological Changes, *Fish Shellfish Immunol.*, (2006), 4, pp. 372-384
- [11] Ohkhawa, H., Ohishi, N., Yogi, K., Assay for Lipid Peroxidation in Animal Tissue by Thiobarbituric Acid Reaction, *Anal Biochem*, (1979), pp. 95-351
- [12] Moron, M. S., Depiere, J. W., Mannervik, B., Levels of GSH, GR and GST Activities in Rat Lung and Liver, *Biochim Biophys Acta.*, (1979), 582, pp. 67-78
- [13] Beutler, E., Duron, O., Kellin, B. M., Improved Method for the Determination of Blood Glutathione, *J Lab Clin Med.*, (1963), 61, pp. 882-888
- [14] Nubel, T., et al., Lovastatin Protects Human Endothelial Cells from Killing by Ionizing Radiation Without Impairing Induction and Repair of DNA Double-Strands Breaks, *Clin Cancer Res.*, (2006), 12, pp. 933-939
- [15] Potton, C. S., A Comprehensive Study of the Radiobiological Response of the Murine (BDF 1) Small Intestine, *Int. J. Radiat Biol.*, (1990), 58, pp. 925-973
- [16] EI-Habit, O. H., et al., The Modifying Effect of b-Carotene on Gamma Radiation: Induced Elevation of Oxidative Reactions and Genotoxicity in Male Rats, *Mutat Res.*, (2000), 25, pp. 551-560
- [17] Singh, I., Soyil, D., Goyal, P. K., Embilca Officinalis (Linn) Fruit Extract Provides Protection Against Radiation-Induced Hematological and Biochemical Alterations in Mice, *J. Environ Pathol Toxicol Oncol.*, (2006), 25, pp. 643-654
- [18] Sancheti, G., Goyal, P. K., Prevention of Radiation-Induced Haematological Alterations by Medicinal Plant Rosemarinus Officinalis in Mice, *Afr. J. Altern Complement Med.*, (2007), 4, pp. 165-172
- [19] Jahan, S., Prophylactic Use of Alstonia Scholaris (Sapthaparna) Against Gamma Irradiation, *Pharmacologyonline*, (2009), 1, pp. 160-175
- [20] Jagetia, G. C., Baliga, M. S., The Evaluation of Nitric Oxide Scavenging Activity of Certain Indian Medicinal Plants in vitro, A Preliminary Study, *J. Med. Food*, (2004), 7, pp. 343-348
- [21] Chaudhary, R., et al., Radioprotective Potential of Trigonella Foenum Graecum Seeds Extract, *Pharmacologyonline*, (2008), 2, pp. 14-26
- [22] Flidner, T. M., Nothdurft, W., Steinbach, K. H., Blood Cell Changes after Radiation Exposure As an Indicator for Hemopoietic Stem Cell Function, *Bone Marrow Transplantation*, (1988), 3, pp. 77-84
- [23] Arora, R., et al., Cytoprotective Effect of Podophyllum Hexandrum against Gamma Radiation is Mediated Via Hemopoietic System Stimulation and Up-Regulation of Heme-Oxygenase-1 and the Prosurvival Multidomain Protection Bcl-2, *Integrative Cancer Therapies*, 6 (2007), 1, pp. 54-65
- [24] Samarth, R. M., Goyal, P. K., Kumar, A., Radioprotective Effects of Mentha Pipertia, *J. Med. Aromatic Plant Sci.*, 22/23 (2001), 4 A/1A, pp. 91-97
- [25] Gridley, D. S., Dose and Dose Rate Effects of Whole: Body Gamma-Irradiation: II. Hematological Variables and Cytokines, *In vivo*, (2001), 15, pp. 209-216
- [26] Tawfik, S. S., Efficiency of Taurine Usage as Treatment for Exposure to Ionizing Radiation Dissertation, Cairo, Egypt: Institute of Environmental Studies and Researches, Ain Shams University, 2003

- [27] Nunia, V., Goyal, P. K., Prevention of Gamma Radiation- Induced Anemia in Mice by Diltiazem, *J. Radiat Res.*, (2004), 45, pp. 11-17
- [28] Nunia, V., Sancheti, G., Goyal, P. K., Protection of Swiss Albino Mice Against Whole-Body Gamma Irradiation by Diltiazem, *Br. J. Radiol.*, (2007), 80, pp. 77-84
- [29] Jagetia, G. C., Venkatesh, P., Baliga, M. S., Evaluation of the Radioprotective Effect of Aegle Marmelos (L.) Correa in Cultured Human Peripheral Blood Lymphocytes Exposed to Different Doses of Gamma Radiation: A Micronucleus Study, *Mutagenesis*, (2003), 18, pp. 387-393
- [30] Kim, H. J., Radioprotective Effect of An Acidic Polysaccharide of Panax Ginseng on Bone Marrow Cells, *J. Vet. Sci.*, (2007), 8, pp. 39-44
- [31] Jagetia, G. C., Venkatesh, P., Inhibition of Radiation-Induced Clastogenicity by Aegle Marmelos (L.) Correa in Mice Bone Marrow Exposed to Different Doses of Gamma-Radiation, *Hum Exp Toxicol*, (2007), 26, pp. 111-124
- [32] Esterbauer, H., Zollner, H., Schaur, R. J., ISI Atlas of Science, *Biochem.*, (1988), 1, pp. 311-317
- [33] Bump, E. A., Brown, J. M., Role of Glutathione in the Radiation Response of Mammalian Cells in vitro, and in vivo, *Pharmacol. Ther.*, 47 (1990), 1, pp. 117-136
- [34] Murray, D., McBride, W. H., Radioprotective Agents, in: Krik-Othmer Encyclopedia of Chemical Technology, 4th ed., John Wiley and Sons, New York, USA, 1996, 20, pp. 963-1006
- [35] Weiss, J. F., et al., Effect of Radioprotective Agents on Survival after Acute Intestinal Radiation Injury, in: Radiation and the Gastrointestinal Tract (Eds. A. Dubois, G. L. King, D. R. Livengood), CRC Press. Boca Raton, Fla., USA, 1995, pp. 183-199
- [36] Petkau, A., Role of Superoxide Dismutase in Modification of Radiation Injury, *Brit. J. Cancer*, 55 (1987), 8, pp. 87-95
- [37] Seifter, E., et al., Morbidity and Mortality Reduction by supplemental Vitamin A or β Carotene in CBA Mice Given Total Body Gamma Radiation, *J. Natl. Can. Inst.*, (1984), 73, pp. 1167-1177
- [38] Kamalakannan, N., Stanely Mainzen Prince, P., Effect of Aegle Marmelos Correa, (Bael) Fruit Extract on Tissue Antioxidants in Streptozotocin Diabetic Rats., *Ind. J. Exp. Biol.*, (2003), 41, pp. 1285-1288
- [39] Nadkarni, K. M., Indian Materia Medica, (Popular Prakashan, Bombay, India), 1992, Vol. 1, pp. 45-82
- [40] Rastogi, R. P., Mehrotra, B. N., Compendium of Indian Medicinal Plants, (Publication and Informations Directorate, New Delhi), 1991, 2, pp. 17-32

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ХЕМАТОЛОШКЕ ПРОМЕНЕ ИНДУКОВАНЕ РАДИЈАЦИЈОМ И ЊИХОВА ИНХИБИЦИЈА ЕКСТРАКТОМ ВОЋА *AEGLE MARMELLOS*

Ова студија спроведена је да осветли радиозаштитни потенцијал екстракта воћа *Aegle Marmelos* од радијацијом изазваних хематолошких и биохемијских промена у крви и јетри мишева. У ову сврху, одрасли швајцарски бели мишеви изложени су гама зрачењу дозе од 6 Gy уз свакодневно аплицирање екстракта од 100 mg по једном килограму телесне тежине животиње (експериментална група), или без апликације екстракта (контролна група). Изложеност зрачењу резултовало је значајним смањењем читаних еритроцита, хемоглобина и хематокрита у периферној крви. Насупрот, озрачене животиње претретиране екстрактом имале су значајан раст у свим овим састојцима крви у поређењу са озраченом контролном групом. Осим тога, забележен је значајан раст преко нормале липидне пероксидације у озраченој контролној групи, док је пораст био знатно мањи у екстрактом претретираним животињама. Такође, претретман са екстрактом изазвао је значајан раст у нивоима глутатиона у серуму, као и у јетри, у поређењу са контролном озраченом групом. Ови резултати указују на то да екстракт може бити одговоран за заштиту матичних ћелија у коштаном сржи, накнадно доводећи до пораста хематолошких конституената у периферној крви. Ова студија потврђује профилактичку употребу *Aegle Marmelos* воћног екстракта против радијацијом изазваних хематолошких и биохемијских промена код сисара.

Кључне речи: гама зрачења, глутатион, хематолошки састојци, липидна пероксидација, *Aegle Marmelos*, швајцарски бели миш